**INTRODUCTION**

Alzheimer’s disease (AD) is a multifactorial disease, complex, and progressive affecting the older population; the most observed dementia cases is in persons over 65 years of age [1, 2].

Histological and pathological studies of AD have revealed that multiple cellular pathways are involved in AD progression [3]. The pathological features identified in the central nervous system (CNS) in AD are amyloid plaques, neurofibrillary tangles, inflammatory processes and disturbance of neurotransmitters [4, 5]. The dysfunction and degeneration of synapses in AD may involve Aβ-induced oxidative stress compromises the mitochondrial function by an oxidative-stress-mediated mechanism [6].

Reactive oxygen species (ROS) are produced by many redox processes that normally occur in the metabolism of aerobic cells. These species are very reactive and harmful to the cells. If not eliminated, ROS can damage vital molecules, such as proteins, DNA, and lipids. Cells express several defense mechanisms, including antioxidant enzymes and non-enzymatic compounds that help prevent the damaging effects of ROS [7].

Oxidative stress can also play an important role in the development of neurodegenerative disorders, such as Alzheimer’s and Parkinson’s diseases [8, 9]. There is an increasing interest in natural antioxidants, namely the phenols, present in medicinal and dietary plants, that might help prevent oxidative damage. [10-14]. In situations of increased free radical generation, the reinforcement of endogenous antioxidants via intake of dietary antioxidants may be of particular importance in attenuating the cumulative effects of oxidatively damaged molecules. In recent years, the consumption of *Hypericum perforatum* (St. John’s wort) derived products have increased dramatically, and presently it is one of the most consumed medicinal plants over the world [15]. The commercially available *H. perforatum* derived products include sophisticated phytopharmaceuticals and nutraceuticals, teas, tinctures, juices and oil macerates [16]. *H. perforatum* has a wide range of medicinal applications, including skin wounds, eczema, burns, diseases of the alimentary tract and psychological disorders. Nowadays, its use in the treatment of mild to moderate depression has become prominent [17-19]. Numerous papers have been published concerning these aspects and several recent reviews can be pointed out [19-21]. In spite of this intense research activity, the antioxidant potential of *H. perforatum* extracts has been somewhat neglected.

Current studies suggested that *H. perforatum* possessed protective effects against H₂O₂-induced apoptosis of PC12 cells than might be useful in the treatment of oxidative stress related to neurodegenerative diseases such as Alzheimer disease [22].

The chemical composition of the *Hypericum* species is composed of naphthodianthrones (especially hypericin and pseudo-hypericin), acyphloroglucinol derivatives (especially hyperforin and adhyperforin), flavonoids (especially queretin, quercitin, hyperoside and bapigenin), tannins, n-alkanes, xanthones and essential oils [18, 23, 24].

The aim of this study was to identify the potential of *H. perforatum* as a protective and therapeutic agent against neurodegenerative disorders and Alzheimer’s disease.

The chemical composition of ethanolic extract of *H. perforatum* was analyzed by using HPLC-DAD. In order to determine the antioxidant potential of *H. perforatum*, was evaluated by employing several antioxidant tests. The cytotoxic activity of the extract was also determined by using MTT cell cytotoxicity screening assay on HeLa and NRK-52E cell lines. The *in vivo* studies on Swiss male mice were determined by utilizing behavioral, memory tests and histological analysis.
This represents the first report on the chemical composition, antioxidant, cytotoxic and anti-Alzheimer activity of *H. perforatum* that has been tested in this study.

**MATERIALS AND METHODS**

**Plant and animals**

Aerial parts of *H. perforatum* were collected in Malatya (Turkey) in June 2010. The plant material was identified by Nihat Kılıç and a voucher specimen (ISTE 93192) was deposited in the Herbarium of the University Of Istanbul Faculty Of Pharmacy, Istanbul, Turkey.

The ethanol extracts were obtained by maceration of the plant material with ethanol for 3 d at room temperature and this procedure currently was repeated twice. The respective extracts were filtered and dried under reduced pressure at a temperature below 218.15 K. The crude methanol extract was lyophilized and stored at 153.15 K.

The *in vivo* studies were performed on Swiss mice (3-month-old), 22-26 g, purchased from Pasteur Institute of Algiers. Mice were housed in the Laboratory animal care of Mostaganem University in 12-hour light-dark cycles at 196.15 K with free access to food and water.

**Chemical agents**

Hypericin, chlorogenic acid, rutin, hyperoside, quercetin, kaempferol, quercetin, amentoflavone, hyperforin, AA, and D-Galactose were obtained from Sigma-Aldrich (Taufkirchen, Germany). Pseudohypericin was trained from PhytoPlan (Heidelberg, Germany). Milk-Q ultrapure water was obtained from Millipore (Billerica, MA). HPLC grade acetonitrile, methanol, ethyl acetate and sodium dihydrogen phosphate dihydrate were obtained from Merck (Darmstadt, Germany) and orthophosphoric acid 85% (Heidelberg, Germany). Milli-Q ultrapure water was obtained from Millipore (Billerica, MA). Dulbecco’s Modified Eagle Medium (DMEM), penicillin, streptomycin, phosphate buffer saline (PBS) were purchased from Roche (Mannheim, Germany). All other reagents were of analytical grade.

**Experiment 1**

The extract has been tested by HPLC-DAD. The HPLC system housed in the Laboratory animal care of Mostaganem University in 12-hour light-dark cycles at 196.15 K with free access to food and water.

**In vitro antioxidant activity**

Total soluble phenolic in the ethanolic extracts was determined with Folin-Ciocalteu reagent according to the method of Slinkard & Singleton with some modifications [27].

The amount of total phenolic compounds was calculated from the calibration curve of Gallic acid standard solution (covering the concentration range between 0.05 and 0.4 mg/ml) and expressed as mg Gallic acid equivalents (GAE)/g dry weight (DW) of the plant material.

Total flavonoid content was established by using a method described by Sakana [28]. Total flavonoid contents were calculated based on the calibration curve prepared with standard catechin solution and expressed mg of (+)-catechin equivalents (CE) per g of DW of the plant material.

The DPPH radical scavenging activity of the methanolic extracts was measured according to the procedure described by Brand-Williams [26]. Two controls were utilized for this test, a negative control (containing all reagents except the test sample) and positive controls (using the reference antioxidants). The ability to scavenge DPPH radical was calculated by the following equation:

\[
\text{AA (DPPH radical scavenging activity, } \% \text{)} = \frac{(1 - \text{Abs sample} / \text{Abs control}) \times 100}{(\text{Abs sample} / \text{Abs control})}
\]

Lipid peroxidation (LPO) assay was prepared on the basis of the method described by Duh et al. and Brand-Williams et al. [39-30]. Formation of LPO products was assayed by the measurement of malondialdehyde (MDA) levels on the basis of MDA reacted with TBA at 532 nm according to Buege & Aust. The percentage inhibition of LPO was calculated by comparing the results of the sample with those of controls not treated with the extract using the following equation:

\[
\text{Inhibition effect, } \% \text{)} = 1 - \frac{(\text{Abs sample} / \text{Abs control})}{(\text{Abs sample} / \text{Abs control})} \times 100
\]

The effect of the ethanolic extract on the generation of superoxide radicals was determined by the nitro blue tetrazolium (NBT) reduction method of Nishikimi [29]. Abilities to scavenge the superoxide radical were calculated by comparing the results of the
sample with those of controls not treated with the extract using the following equation:

\[ \text{SOD scavenging (\%)} = \frac{1}{1 - \frac{\text{Abs sample (560nm)}}{\text{Abs control (560nm)}}} \times 100 \]

FRAP assay was carried out in accordance with the procedure of Benzie & Strain [31-33]. The standard curve was constructed using iron sulphate heptahydrate solution (125-2000 µM), and the results were reported in mm Fe²⁺-equivalents.

**In vitro cytotoxic activity**

Cytotoxicity of the extract at various concentrations was determined on human cervix adenocarcinoma (HeLa, ATCC® CCL-2®) and normal rat kidney epithelial (NRK-52E, ATCC® CRL-6509™) cell lines by the MTT assay, which is widely used for the measurement of cell viability. Briefly, the cells were seeded in 96-well plates at a density of 104 cells/well in 100 µl culture medium. Following 24-h incubation and attachment, the cells were treated with different concentrations of plant extracts and controls for 24 h. Dry methanolic extracts were dissolved in dimethyl sulphoxide (DMSO) as a solvent to obtain appropriate stock solutions of the extracts. Dilution of stock extracts of solutions was made in serum-free medium yielding final extracts concentrations from 0.125 to 2 mg/ml. DMSO and 5-fluorouracil (5-FU) were used as solvent and medium yielding final extracts concentrations from 0.125 to 2 µM. The yellow MTT dye was reduced by succinic dehydrogenase in the mitochondria of viable cells to purple formazan crystals. The absorbance was measured by a microplate reader (BioTek, USA) at 570 nm with a reference wavelength of 670 nm. The reduction of absorbance was evaluated the inhibition of enzyme activity observed in cells compared to untreated (negative control) cells. Then, the half maximal inhibitory concentration (IC50) was expressed as the concentration of the sample that caused an inhibition of 50% in enzyme activities in cells as flows [34].

\[ \text{IC} 50 = \frac{1}{[\text{mean absorbance of extract} \times 100]} \]

The results were generated from three independent experiments; each experiment was performed in triplicate.

**Experiment 3**

**In vivo experimental design included three groups of mice**

Control group: Mice were administered freshwater orally and served as normal control. Alzheimer's model group: Mice were treated with AlCl₃ (100 mg/kg/day) concomitant with IP D-Gal in order of 1200 s divides to four phases, 300 s each. The LC chromatograms of the ethanolic extract of H. perforatum

**Behavioral and memory tests**

The different groups were monitored for the onset of postural abnormalities, general behavioral changes memory and learning.

Locomotor activity refers to the movement from one location to another. In rodents, one of the most important components of exploration, a prominent activity of the rat’s repertoire of impulsive activity, is locomotion. We detected spontaneous motor activity over a four periods of 1200 s (300 s each) in an activity cage. Measure of motor exercise endurance was evaluated using a divided box apparatus counting the number of visited cases.

The anxiety test was established by means of dark/white connected rooms. The light/dark test is built on the innate aversion of rodents to brightly illuminated areas and on the spontaneous exploratory behavior of rodents in response to mild stressors, that is, novel environment and light. The test apparatus consists of a small obscure safe compartment (half box) and a large floodlit aversive compartment. Mice are placed in one of the rooms and the time spent in each room was assessed. These tests were realized during 1200 s divides to four phases, 300 s each.

The hole board test was used to assess the behavior of mice confronted with a new environment. The test is to determine the effects of psychotropic drugs on exploratory behavior exhibited by the animal. It only takes a few minutes and does not require learning or conditioning of animals from them. Exploration holes movements are counted as a score of exploration.

The study of memory incorporates research methodologies from neuropsychology, human development and animal testing using a wide range of species.

Understanding memory has benefited greatly from animal research. Animal’s brains can be selectively lesions using neurotoxic methods (AlCl₃ in this case) and be assessed comparatively to the treated mice. The assessment of animal memory utilizing different types of mazes has been largely used in neurosciences. Several models have been proposed recently, mainly trying to evaluate the accuracy of choice between the alternatives presented in the same day of the session, instead of looking for the accumulated learning through successive days of training.

Non-spatial memory was used in neurology in order to assess memory disorder depends upon the hippocampus. For this aim of the experiment, only two arms of the maze were utilized. A baited lighted arm and the other arm were unlit. The subject is well placed on the central platform with two closed arms, and the two arms are straight successively for it to adapt to its new environment. The spending time in the lighted arm was assessed.

Working memory relates to a short-term memory system used to complete a task, such as a relationship between objects and locations. To demonstrate that mice were matched for memory ability after the treatment period, working memory was evaluated in averaging the reentries and error entries, respectively, for the successive days of experimentation.

**Histological study**

Histological, analyses were performed on formalin-fixed and paraffin-embedded brain tissues from the experiment’s treatment groups. Briefly, whole formalin-fixed brains were cut in 2 mm thick slices along the sagittal plane and embedded in formalin. 2 mm thick serial sections were stained with Haematoxylin-Eosin, periodic acid Schiff, Luxol fast blue and Perls’ stain.

**Statistical analysis**

About in vivo studies, the data are expressed as means with SEM. The statistical significance of differences between groups was assessed with an analysis of variance followed by Student Newman-Keuls. A P value of 0.05 or less was considered as a criterion for a statistically significant difference.

**RESULTS**

**Experiment 1**

The LC chromatograms of the ethanolic extract of H. perforatum were showed in figure 1.
Fig. 1: It shows the LC chromatogram of ethanolic extract of *H. perforatum* A: Pseudohypericin (1), hypericin (2) in the extract at 590 nm (fig. 1A) B: Chlorogenic acid (1), rutin (2), hyperoside (3), isoquercitrin (4), quercitrin (5), kaempferol (6), quercetin (7), amentoflavone (8) in the extract at 360 nm (fig. 1B) C: Hyperforin (1) in the extract at 275 nm (fig. 1C)

**Experiment 2**

*In vitro* antioxidant activity

In table 2 and table 3.

**In vitro cytotoxic activity**

Ethanolic extract of *H. perforatum* was evaluated for *in vitro* cytotoxic activity using the colorimetric MTT assay in HeLa and NRK-52E cell lines. The IC50 value (mg/cm²) is given in the table. The extract did not demonstrate significant cytotoxic activity against HeLa and NRK-52E cell lines.

**Experiment 3: *In vivo* study**

Behavioral tests

The performance of mice in the step-through passive avoidance training and testing shows a remarkable difference between Alzheimer treated, and the Alzheimer’s model groups; locomotors activity test showed a hyperactivity in Alzheimer’s model group comparatively to the treated Alzheimer’s and control groups.

Anxiety tests presented in this study with the dark and light rooms, showed significant results for the treated Alzheimer’s comparatively to the Alzheimer’s model group.

The test of maze plus was assessed by the time spending in a covered arm squares noted as score per time of 1200 s, 300 s each phases showed high activity for the Alzheimer’s treated comparatively with the Alzheimer’s model group. fig. 2.

**Memory tests**

Non-spatial memory test preferably conditional results obtained during the five experimental tests show that Alzheimer’s model mice spent shortly time in the arm lit unlike treated Alzheimer’s and control mice, that put a very short time to get informed on the arm.

For the work spatial memory, the results of the treated Alzheimer’s group was very significant and very close to the control group results in the last experiment day comparatively to the Alzheimer’s model group. fig. 3.

**Table 1: it shows the chemical compounds of the extract found in HPLC chromatogram**

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Retention time (min)</th>
<th>Calibration equation values</th>
<th>Linear regression (r²)</th>
<th><em>H. perforatum</em> (yield, %)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pseudohypericin</td>
<td>4.86</td>
<td>y = 2.582269e+007x+1741.874</td>
<td>0.9998</td>
<td>0.075±0.0100</td>
</tr>
<tr>
<td>Hypericin</td>
<td>13.93</td>
<td>y = 6.03411e+007x+297.2292</td>
<td>0.9999</td>
<td>0.067±0.0400</td>
</tr>
<tr>
<td>Chlorogenic acid</td>
<td>4.33</td>
<td>y = 5110294x+1490.398</td>
<td>0.9999</td>
<td>0.008±0.0010</td>
</tr>
<tr>
<td>Rutin</td>
<td>8.89</td>
<td>y = 1.38350e+007+5181.852</td>
<td>0.9999</td>
<td>1.015±0.0400</td>
</tr>
<tr>
<td>Hyperoside</td>
<td>10.19</td>
<td>y = 2.84991e+007x+526.7023</td>
<td>0.9999</td>
<td>0.491±0.0900</td>
</tr>
<tr>
<td>Isoquercitrin</td>
<td>10.75</td>
<td>y = 1.67113e+007x-372.788</td>
<td>0.9999</td>
<td>0.389±0.0600</td>
</tr>
<tr>
<td>Quercitrin</td>
<td>14.41</td>
<td>y = 1.205178e+007-3518.974</td>
<td>0.9999</td>
<td>0.135±0.0100</td>
</tr>
<tr>
<td>Kaempferol</td>
<td>17.09</td>
<td>y = 5.18391e+007x+437.856</td>
<td>0.9999</td>
<td>0.002±0.0001</td>
</tr>
<tr>
<td>Quercetin</td>
<td>17.84</td>
<td>y = 3.688175e+007+1890.53</td>
<td>0.9999</td>
<td>0.125±0.0500</td>
</tr>
<tr>
<td>Amentoflavone</td>
<td>20.27</td>
<td>y = 2.207879e+007+72.0972</td>
<td>0.9996</td>
<td>0.06±0.0002</td>
</tr>
<tr>
<td>Hyperforin</td>
<td>27.75</td>
<td>y = 6212343x</td>
<td>0.9997</td>
<td>3.375±0.5000</td>
</tr>
</tbody>
</table>

**Table 2: it shows the total extractable compounds (EC), total phenolic compounds (PC) (As Gallic acid equivalents) and total flavonoids (as catechin equivalents) in the extract**

<table>
<thead>
<tr>
<th>Extracts</th>
<th>EC (mg/g DW*)</th>
<th>PC (mg/g DW*)</th>
<th>Flavonoid (mg/g DW*)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>H. perforatum</em></td>
<td>267.8</td>
<td>34.7 ± 0.9</td>
<td>31.8 ± 0.9</td>
</tr>
<tr>
<td>DW: Dried Weight</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Table 3: it shows the antioxidant activity (EC50 values)**

<table>
<thead>
<tr>
<th>Extracts</th>
<th>Average of EC50 (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Anti-LPO</td>
</tr>
<tr>
<td><em>H. perforatum</em></td>
<td>2.74 ± 0.19</td>
</tr>
<tr>
<td>Quercetin</td>
<td>0.059 ± 0.001</td>
</tr>
</tbody>
</table>

*Values were the means of three replicates ± standard deviation.*
The cytoplasm of neuron were shrunken, the nuclei were side moved and nuclei were dark stained, neurofibrillary degeneration and neuron loss were observed in the hippocampus. *Hypericum perforatum* administration showed moderated neuropathological changes. The treatment decreased the shape modifications of the neurons, with prolonged neurofibrillary reactions. fig. 4.

**DISCUSSION**

A complex mixture of bioactive secondary metabolites in several *Hypericum* species makes them valuable as herbal drugs [35-37]. *H. perforatum* (Common St. John’s wort), certainly the best known and worldwide the most abundant representative, is today the most investigated species of the genus [38]. Since the early 90’s, *H. perforatum* has been clinically studied from the perspective both of its chemical constituency and of its biological activity [39]. Around 2500 studies on *Hypericum* have been published to date (thereof c. 950 without *H. perforatum*; S. Crockett, pers. com.), including several reviews focused on the phytochemistry of *H. perforatum* [38,40-41], its pharmacology [42,44] or both aspects [38,45-47].

Phytochemical analysis of *H. perforatum* was achieved for the first time in this work; it was investigated the medicinal value of *H. perforatum*. According to European Pharmacopoeia, *H. perforatum* is a traditional medicinal plant with antidepressive and antioxidant properties. It contains a large number of constituents with documented biological activity including phenolic, a broad range of flavonoids and bioflavonoids, hypericin, adhyperforin, amentoflavone, hyperoside and isoquercitrin [47]. The chemical compounds of *H. perforatum* are provided in table 1. A total of 11 components were defined in the extract. Hyperforin constituted the major fraction (3.375%) this is due to the fact that hyperforin is photosensitive and its highest concentrations appear just before the blossom period [48].

It has been determined that hyperforin, a major constituent of *H. perforatum*, inhibits the proliferation of epithelial cells and human peripheral blood mononuclear cells, even when stimulated by phyto-hemagglutinin. Furthermore, another study [49], also justified the anticancer activity of extracts of *H. perforatum* by the presence of hyperforin and hypericin of the plant. Previous work [50] De Freitas Silva et al. demonstrated that compounds other than hypericin are highly relevant for both radical scavenging and inhibition of lipid peroxidation activities.

Rutin constituted the second major fraction, accounting for 1.015% of the extract. Umek et al. found that the content of rutin in *H. perforatum* was in a strong positive correlation with the altitude of the growing site, opposite to the quercitin content [51].

Chlorogenic acid has been reported to inhibit lipid peroxidation by scavenging peroxyl radicals, thereby preventing the initiation of chain lipid peroxidation [50]. Also, chlorogenic acid had been reported to be an effective scavenger against peroxyl radicals [50]. According to the previous research [52], Chlorogenic acid has an effective antioxidant activity. These data are in accordance with the recently published chemical characterization of *H. perforatum* of Macedonia [48].

In the present study, using various in vitro experiments, we found that *H. perforatum* markedly inhibited XO activity, suggesting that besides direct scavenging [53,54]; inhibition of the XO enzyme may contribute to the scavenging action of the O2-radical by *H. perforatum*.

Considering antioxidant activity of the marine sponge species screened herein, the literature survey indicated that there has been only one former study describing the antioxidant potential of the compounds; 2-octaprenyl-1,4-hydroquinone and 2-(24-hydroxy)-octaprenyl-1,4-hydroquinone isolated from *I. spinosula* and its eight synthetic derivatives [55]. Their antioxidant activity was tested in *vitro* by scavenging of free radical DPPH and inhibition of the lipid peroxidation induced by the Fe++/ascorbate system. Two natural hydroquinone derivatives were found to display high antioxidant effect in both tests, which was in accordance with our results. In our DPPH radical scavenging assay, the highest scavenging effect (65.92%) was observed in 1. variability of Antalya-I collection (table 3) and linear terpenes including.

**Table 4: It shows the cytotoxic potential of the extract (IC50 value)**

<table>
<thead>
<tr>
<th>Extracts</th>
<th>IC50 values (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HeLa</td>
<td>0.541</td>
</tr>
<tr>
<td>NRR-52E</td>
<td>0.619</td>
</tr>
</tbody>
</table>

*Positive Control: (5-FU): 48.012 µM for HeLa, 12.645 µM for NRR-52E

Histological studies

H&E staining shows that there are typical neuropathological changes in the cerebral cortex and hippocampus of Alzheimer’s model. In the control groups, the neurons were full and arranged tightly; the nuclei were lit stained. By comparison in the model group mice the cytoplasm of neuron were shrunken, the nuclei were side moved and dark stained, neurofibrillary degeneration and neuron loss were observed in the hippocampus.
The extract showed the DPPH radical scavenging activity in a dose-dependent manner. The highest antioxidant activity of the extract was observed in high concentration (5 mg/cm³). The antioxidant activity of the extract was determined lower than quercetin activity. Based on the EC₅₀ values, the antioxidant activity of HPM was significantly lower (p<0.05) with an EC₅₀ value of 2.74 ± 0.219 mg/cm³ in comparison with quercetin (0.059 ± 0.001 mg/cm³).

Although the inhibitory effect of the extract on TBARS formation was less than that of quercetin, the extract may also be supposed to protect against damage to cell membranes, because it also reduces the level of peroxides. However, the extract has the capacity to remove free radicals and inhibit lipid peroxidation. The results consistent with the other in vitro studies with Hypericum species. Couladis and Silva et al. have reported that the methanolic extract of H. perforatum shown to inhibit lipid peroxidation with an EC₅₀ value of 26 µg/cm³ [59,60]. It is reported that the methanolic extract of H. perforatum shown to inhibit (70%) lipid peroxidation [57]. It was also reported by Kızıl et al. that the extracts obtained from H. triquetrifolium and H. scabroides showed high potential to inhibit lipid peroxidation [56].

The extract was found to be an efficient scavenger of superoxide radical generated in PMS/NADH system, and its superior activity was comparable (p>0.05) to that of quercetin. Both extract and quercetin exhibited dose-dependent inhibition of superoxide radical.

The reducing power of the extract was found to be correlated with increasing absorbance (at 700 nm) as compared with quercetin. The extract has shown the potential of reducing ferric (III) form of iron to ferrous (II). These results compared with previous studies, similar results were reported for H. triquetrifolium and H. scabroides [56].

Antioxidant activity of the extract was examined by using four different methods. Table 3 shows EC₅₀ values of the extract in antioxidant properties. The differences of the results explained diverse mechanisms of oxidative stress.

There was a great correlation between total phenolic contents and reductive potential and lipid peroxidation inhibitory activity. As a result, this extract has antioxidant activity due to the phenolic compounds have the potential to act as antioxidants by scavenging free radicals, reducing power and blocking the formation of peroxide radicals and reducing power.

Since the cytotoxic activity potential of the extracts having antioxidant activity is an important parameter, the cytotoxic activity of the HeLa and NRK-52E cell lines is investigated. The results reveal that the extracts have no cytotoxic activity.

Antidepressant activity of H. perforatum is certainly the main reason for the general public's enthusiasm for this herbal medication [43]. The classical use of (dried alcoholic) extracts of H. perforatum for the treatment of mild to moderate depression has been demonstrated to be effective in several trials and meta-analyses, in the same citation the antidepressant activity of H. perforatum-based formulations can be attributed to several classes of secondary metabolites, which...
exhibit additive, synergetic and partly antagonistic effects [61,42]. According to the actual state of scientific knowledge, the total extracts must be considered as the active principle [41].

The effects of H. perforatum on the central nervous system were investigated using various behavioral and memory models, including locomotors activity, dark/lighted rooms and hole-board for the behavior assessment, non-spatial memory and worked spatial memory for the memory experiments. According to the results, it was found that extracts prepared from H. perforatum were effective as anti-depressant drug. This conclusion suggested that the anti-depressant effects of H. perforatum were remarkable in behavioral tests and may be used for therapeutic purposes in depression [62]. The species [rodents] in which aluminum-induced neurobehavioral effects (e.g., changes in locomotors activity, learning and memory) have been observed fail to develop significant cytotoxicological pathology, but exhibit a number of neurochemical alterations following in vivo or in vitro exposure [63,64]. In this case, the locomotors activity decreased in treated Alzheimer’s comparatively to the Alzheimer group that confirms that hyperactivity observed for the Alzheimer’s group could be the result of stress conditions [65].

Anxiety-related behavior is measured as a preference for the closed arms [66-68]. The percentage of open arm entry also indicates anxiety levels, especially in mice, which tend to be more impulsive and spend less time per entry in any arm. In this study, the time spent in opens area and close to the control group. The hole-board test provides a simple method for measuring the response of an animal to an unfamiliar environment and is widely used to assess emotionality, anxiety and/or responses to stress [69-71].

After the treatment administration of both Aluminum and Hypericum the head dip counts decreased without changing in locomotion. These results indicate that H perforatum has satisfactory in axiolytic effect in this paradigm [72]. After “training” an experimental animal, such as a mouse, the only way to be sure that a “memory” was formed is by evoking it back, i.e., by recalling it in a “test” session: this “memory” is expressed by a behavior that differs from that one emitted in the training session. Training sessions consist of repeating a number of trials, several days in a row [4-8 trials a day, for 2 to 5 d or more, when training to a criterion] [73]. Non-spatial memory conditional preference showed an improvement from the third day as same as the work spatial memory that improved from the 4th day, that result confirm the damage reduction in the brain in treated Alzheimer’s comparatively to the Alzheimer group.

More specifically, the entire brain of patients with Alzheimer’s disease (AD) was shown to be subjected to an oxidative challenge [74]. In addition, overall peroxidation activity in brains of AD patients was significantly elevated compared to controls [75]. In the reverse phenomena, H. perforatum extract contains flavonoids such as rutin, quercitin, and quercitrin, which demonstrated a free radical scavenging activity in a model of auto-oxidation of rat cerebral membranes [76].

In this investigation, the effect of H. perforatum lead to a reduction of neurotoxicity and Alzheimer’s disease appeared as shrunken decreased in pyramidal cells, reduced effect of the decreasing number of the pyramidal cells. These brain moderation changes inducing by H. perforatum Alzheimer’s were due to reducing oxidative damage and edema which contribute to disease pathogenesis and were in accordance with some researchers [77].

CONCLUSION

Operative treatments and medications are still absent to seize Alzheimer’s diseases. AD is a progressive neurodegenerative disorder that affects the elderly population particularly. Since pathogenesis of AD has not been clarified totally, yet, it is only a symptomatic treatment available. The most prescribed drug class against AD is cholinesterase inhibitors, which increase the level of acetylcholine/butyryl-choline in the brain.

Another factor contributing to the pathology of neurodegenerative diseases is oxidative stress, which leads to neuronal death. Inflammation is also linked to neurodegeneration by diverse mechanisms such as accretion of proteins with abnormal conformations or via signals emanating from injured neurons.

Considering the herbal treatment, we initiated an intensive research on the neuroprotective activity of the H. perforatum through the enzyme inhibition linked to neurodegeneration and antioxidant activity methods, which have so far let us to realize the in vitro and in vivo studies. Antioxidant activity of the samples has been tested in a number of tests such as 2,2-diphenyl-1-picrylhydrazyl (DPPH), NN-dimethylphenylenediamine (DMPD), superoxide, nitric oxide, hydrogen peroxide104 radical scavenging, ferric-reducing antioxidant power (FRAP), and phospho-molybdenum reducing antioxidant power (PRAP) assays and confirmed by in vivo study showing a decrease in cell degeneration.

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ABBREVIATION

AD: Alzheimer disease; ROS, Reactive oxygen species; HPLC-DAD, High performance liquid chromatography-diode array detector); MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NAD: Nicotinamide adenine dehydrogenase; BHT, Butylated-hydroxytoluene; PMS, Phenazinemethosulphate; EDTA, Ethylenediamine extra acetic acid; TCA, Trichloroacetic acid; DMSO, Dimethylsulfoxide; EDTA, Ethylenediamine tetraacetic acid; FBS, Fetal bovine serum; DMEM, Dulbecco’s Modified Eagle Medium; PBS, phosphate buffer saline; LDH, Lactate deshydrogenase; GAE, Gal acid equivalent.

CONFLICT OF INTERESTS

Declared none

REFERENCES


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